 Estradiol levels have been reported to be increased in patients with feminizing syndromes, gynecomastia, and testicular tumors.

In cases of infertility, serum estradiol measurements are useful for monitoring induction of ovulation following treatment with, for example, clomiphene citrate, LH-releasing hormone (LH-RH), or exogenous gonadotropins. During ovarian hyperstimulation for in vitro fertilization (IVF), serum estradiol concentrations are usually monitored daily for optimal timing of human chorionic gonadotropin (hCG) administration and oocyte collection.

Oxis’s Estradiol (E2) EIA kits are designed for the measurement of total Estradiol in human serum or plasma.

**PRINCIPLE OF THE TEST**

The Oxis E2 EIA is based on the principle of competitive binding between E2 in the test specimen and E2-HRP conjugate for a constant amount of rabbit anti-Estradiol. In the incubation, goat anti-rabbit IgG-coated wells are incubated with 25 µl E2 standards, controls, patient samples, 100 µl Estradiol-HRP Conjugate Reagent and 50 µl rabbit anti-Estradiol reagent at room temperature (18-25°C) for 90 minutes. During the incubation, a fixed amount of HRP-labeled E2 competes with the endogenous E2 in the standard, sample, or quality control serum for a fixed number of binding sites of the specific E2 antibody. Thus, the amount of E2 peroxidase conjugate immunologically bound to the well progressively decreases as the concentration of E2 in the specimen increases. Unbound E2 peroxidase conjugate is then removed and the wells washed. Next, a solution of TMB Reagent is added and incubated at room temperature for 20 minutes, resulting in the development of blue color. The color development is stopped with the addition of 1N HCl, and the absorbance is measured spectrophotometrically at 450 nm. The intensity of the color formed is proportional to the amount of enzyme present and is inversely related to the amount of unlabeled E2 in the sample. A standard curve is obtained by plotting the concentration of the standard versus the absorbance. The E2 concentration of the specimens and controls run concurrently with the standards can be calculated from the standard curve.

**REAGENTS**

- **Materials provided with the kit:**
  - Goat Anti-Rabbit IgG-coated microtiter wells, 96 wells
  - Estradiol Reference Standards: 0, 10, 30, 100, 300, and 1000 pg/ml. Liquid, 0.5 ml each, ready to use.
  - Rabbit Anti-Estradiol Reagent (pink color), 7 ml
  - Estradiol-HRP Conjugate Reagent (blue color), 12 ml
  - Estradiol Control 1, Liquid, 0.5 ml, Ready to use.
  - Estradiol Control 2, Liquid, 0.5 ml, Ready to use.
  - TMB Reagent (One-Step), 11 ml.
  - Stop Solution (1N HCl), 11 ml.

- **Materials required but not provided:**
• Precision pipettes: 25 µl, 50 µl, 100 µl, 200 µl, and 1.0 ml.
• Disposable pipette tips.
• Distilled and deionized water.
• Vortex mixer or equivalent.
• Absorbent paper or paper towel.
• Linear-linear graph paper.
• Microtiter plate reader.

**WARNINGS AND PRECAUTIONS FOR USERS**

Test methods are not available which can offer complete assurance that Hepatitis B virus, Human Immunodeficiency Virus (HIV/HTLV-III/LAV), or other infectious agents are absent from the reagents in this kit. Therefore, all human blood products, including patient samples, should be considered potentially infectious. Handling and disposal should be in accordance with the procedures defined by an appropriate national biohazard safety guideline or regulation, where it exists (e.g., USA Center for Disease Control/National Institute of Health Manual, “Biosafety in Microbiological and Biomedical Laboratories,” 1984)(22).

**SPECIMEN COLLECTION AND PREPARATION**

1. Serum or EDTA plasma should be used.
2. No special pretreatment of sample is necessary.
3. Serum or plasma samples may be stored at 2-8°C for up to 24 hours, and should be frozen at −10°C or lower for longer periods. Do not use grossly hemolyzed or grossly lipemic specimens.
4. Please note: Samples containing sodium azide should not be used in the assay.

**STORAGE OF TEST KIT AND INSTRUMENTATION**

Unopened test kits should be stored at 2-8°C upon receipt and the microtiter plate should be kept in a sealed bag with desiccants to minimize exposure to damp air. Opened test kits will remain stable until the expiration date shown, provided it is stored as described above. A microtiter plate reader with a bandwidth of 10 nm or less and an optical density range of 0-3 O.D. at 450 nm wavelength is acceptable for use in absorbance measurement.

**REAGENT PREPARATION**

1. All reagents should be brought to room temperature (18-25°C) before use.
2. Samples with expected Estradiol concentrations over 1000 ng/ml may be quantitated by dilution with diluent available from Oxis International, Inc.

**ASSAY PROCEDURE FOR SERUM AND PLASMA**

1. Secure the desired number of coated wells in the holder.
2. Dispense 25 µl of standards, specimens and controls into appropriate wells.
3. Dispense 100 µl of Estradiol-HRP Conjugate Reagent into each well.
4. Dispense 50 µl of rabbit anti-Estradiol(E2) reagent to each well.
5. **Thoroughly mix for 30 seconds. It is very important to mix them completely.**
6. Incubate at room temperature (18-25°C) for 90 minutes.
7. Rinse and flick the microwells 5 times with distilled or deionized water. (**Please do not use tap water.)**
8. Dispense 100 µl of TMB Reagent into each well. Gently mix for 10 seconds.
9. Incubate at room temperature (18-25°C) for 20 minutes.
10. Stop the reaction by adding 100 µl of Stop Solution to each well.
11. Gently mix 30 seconds. **It is important to make sure that all the blue color changes to yellow color completely.**
12. Read absorbance at 450 nm with a microtiter well reader **within 15 minutes.**

**CALCULATION OF RESULTS**

1. Calculate the mean absorbance value (A450) for each set of reference standards, controls and samples.
2. Construct a standard curve by plotting the mean absorbance obtained for each reference standard against its concentration in pg/ml on a linear-linear graph paper, with absorbance values on the vertical or Y axis, and concentrations on the horizontal or X axis.
3. Use the mean absorbance values for each specimen to determine the corresponding concentration of Estradiol in pg/ml from the standard curve.
4. Any values obtained for diluted samples must be further converted by applying the appropriate dilution factor in the calculations.

**EXAMPLE OF STANDARD CURVE**

Results of a typical standard run with optical density readings at 450 nm shown in the Y axis against Estradiol concentrations shown in the X axis. **Note:** This standard curve is for the purpose of illustration only, and should not be used to calculate unknowns. Each laboratory must provide its own data and standard curve in each experiment.

<table>
<thead>
<tr>
<th>Estradiol (pg/ml)</th>
<th>Absorbance (450 nm)</th>
</tr>
</thead>
<tbody>
<tr>
<td>0</td>
<td>2.943</td>
</tr>
<tr>
<td>10</td>
<td>2.551</td>
</tr>
<tr>
<td>30</td>
<td>2.055</td>
</tr>
<tr>
<td>100</td>
<td>1.624</td>
</tr>
<tr>
<td>300</td>
<td>0.925</td>
</tr>
<tr>
<td>1000</td>
<td>0.571</td>
</tr>
</tbody>
</table>
EXPECTED VALUES AND SENSITIVITY

Each laboratory should establish its own normal range based on the patient population. Oxis Estradiol EIA was performed on randomly selected outpatient clinical laboratory samples. The results of these determinations are as follows:

Males:          < 60 pg/ml
Females:  postmenopausal phase        < 18 pg/ml
              ovulating, early follicular 30-100 pg/ml
              late follicular 100-400 pg/ml
              luteal phase 60-150 pg/ml
              pregnant, normal up to 35,000 pg/ml
              prepubertal children, normal < 10 pg/ml

PERFORMANCE CHARACTERISTICS

1. Sensitivity

The minimum detectable concentration of the Oxis Estradiol ELISA assay as measured by 2 SD from the mean of a zero standard is estimated to be 10 pg/ml.

2. Precision

   a. Intra-Assay Precision
   Within-run precision was determined by replicate determinations of four different serum samples in one assay. Within-assay variability is shown below:

<table>
<thead>
<tr>
<th>Samples</th>
<th>1</th>
<th>2</th>
<th>3</th>
<th>4</th>
</tr>
</thead>
<tbody>
<tr>
<td># Replicates</td>
<td>24</td>
<td>24</td>
<td>24</td>
<td>24</td>
</tr>
<tr>
<td>Mean Estradiol (pg/ml)</td>
<td>13</td>
<td>73</td>
<td>247</td>
<td>633</td>
</tr>
<tr>
<td>S.D.</td>
<td>3</td>
<td>7</td>
<td>10</td>
<td>31</td>
</tr>
<tr>
<td>C.V. (%)</td>
<td>24.1</td>
<td>10.3</td>
<td>4.1</td>
<td>4.9</td>
</tr>
</tbody>
</table>

   b. Inter-Assay Precision
   Between-run precision was determined by replicate measurements of six different serum samples over a series of individually calibrated assays. Between-assay variability is shown below:

<table>
<thead>
<tr>
<th>Samples</th>
<th>1</th>
<th>2</th>
<th>3</th>
<th>4</th>
</tr>
</thead>
<tbody>
<tr>
<td>Mean Estradiol (pg/ml)</td>
<td>14</td>
<td>82</td>
<td>264</td>
<td>691</td>
</tr>
<tr>
<td>S.D.</td>
<td>4</td>
<td>8</td>
<td>17</td>
<td>45</td>
</tr>
<tr>
<td>C.V. (%)</td>
<td>26.7</td>
<td>10.3</td>
<td>6.4</td>
<td>6.6</td>
</tr>
</tbody>
</table>

3. Recovery Studies

Various patient serum samples of known Estradiol levels were combined and assayed in duplicate. The mean recovery was 101.3%.

4. Specificity

The following materials have been checked for cross reactivity. The percentage indicates cross reactivity at 50% displacement compared to Estradiol.

Data on the cross-reactivity for several endogenous and pharmaceutical steroids are summarized in the following table:

Cross-reactivity (%) = \( \frac{\text{Observed Estradiol Concentration}}{\text{Steroid Concentration}} \) \times 100

<table>
<thead>
<tr>
<th>Steroid</th>
<th>Cross-Reactivity</th>
</tr>
</thead>
<tbody>
<tr>
<td>Estradiol</td>
<td>100%</td>
</tr>
<tr>
<td>Estrone</td>
<td>2.10%</td>
</tr>
<tr>
<td>Estriol</td>
<td>1.50%</td>
</tr>
<tr>
<td>17a Estradiol</td>
<td>0.30%</td>
</tr>
<tr>
<td>Cortisol</td>
<td>&lt;0.01%</td>
</tr>
<tr>
<td>Cortisone</td>
<td>&lt;0.01%</td>
</tr>
<tr>
<td>Progesterone</td>
<td>&lt;0.01%</td>
</tr>
<tr>
<td>Testosterone</td>
<td>&lt;0.01%</td>
</tr>
<tr>
<td>DHEA-Sulphate</td>
<td>&lt;0.01%</td>
</tr>
<tr>
<td>5a-Dihydrotestosterone</td>
<td>&lt;0.01%</td>
</tr>
</tbody>
</table>

CLINICAL APPLICATION

Information is cited from reference #19

1. Assessment of Female Menstrual Dysfunctions:

   - Hyperestrogenism in girls:
     Elevated E2 can be used in the evaluation of precocious puberty in girls. However, extensive ancillary aids are required for specific diagnoses.
   - Hypoestrogenism in women:
E₂ measurements are frequently utilized in the assessment of hypoestrogenism in cases of delayed puberty, primary and secondary amenorrhea, and menopause. In hypoestrogenism women, E₂ concentrations are usually <30 pg/ml.

2. Assessment of Excessive Estrogen Production in Women:
   In pregnant women, E₂ concentrations will >1,000 pg/ml. In non-pregnant women, excessive estrogen may indicate ovarian neoplasms.

3. Monitoring Ovulation:
   E₂ is often measured to monitor ovulation induction and for patient follow-up during infertility therapy, e.g. in vitro fertilization (IVF).

4. Estradiol Measurement in Male:
   E₂ measurement is used in the differential diagnosis of gynecomastia, feminizing syndromes, hypogonadism and testicular tumors.

LIMITATIONS OF THE PROCEDURE
1. Reliable and reproducible results will be obtained when the assay procedure is carried out with a complete understanding of the package insert instructions and with adherence to good laboratory practice.
2. The wash procedure is critical. Insufficient washing will result in poor precision and falsely elevated absorbance readings.
3. Serum samples demonstrating gross lipemia, gross hemolysis, or turbidity should not be used with this test.
4. The results obtained from the use of this kit should be used only as an adjunct to other diagnostic procedures and information available to the physician.

QUALITY CONTROL
Good laboratory practice requires that controls are run with each calibration curve. A statistically significant number of controls should be assayed to establish mean values and acceptable ranges to assure proper performance.

We recommend using Bio-Rad Lyphochek Immunoassay Control Sera as controls. Oxis Estradiol EIA kit also provides with internal controls, Level 1 and 2.

REFERENCES

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